

1 WATER, AIR AND SOIL POLLUTION

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3 **Effects of single, binary and quinary mixtures of phenanthrene and its N-PAHs on *Eisenia fetida* in soil**

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23 **Abstract** It is now acknowledged that aromatic hydrocarbons present in contaminated soils occur in mixtures.
24 The effect of single, binary and quinary mixtures of phenanthrene and selected N-PAHs were investigated on
25 the survival, growth and behavioural index of earthworms (*Eisenia fetida*) over a 21 d incubation in soil. The
26 results showed that the LC₅₀ values ranged from (not detected) ND - 329.3 mg kg⁻¹ (single mixture), ND – 219.8
27 mg kg⁻¹ (binary mixtures) and 148.4 mg kg⁻¹ (quinary mixture), while the EC₅₀ values (based on weight loss)
28 ranged from 13.3 - 148.4 mg kg⁻¹ (single mixture), 63.8 - 148.4 mg kg⁻¹ (binary mixture) and 24.2 mg kg⁻¹
29 (quinary mixture). Greater impacts were recorded where N-PAHs are present with phenanthrene. Further,
30 behavioural index of *E. fetida* was affected after 24 h exposure to N-PAH amended soils. Among the N-PAHs
31 however, benzo[h]quinoline recorded the greatest impact on the survival, growth and behavioural index of *E.*
32 *fetida* in soil. Findings from this study showed that 3 ring-N-PAHs are more toxic than phenanthrene as
33 expected from their physico-chemical properties. Binary and quinary mixtures of phenanthrene and N-PAHs in
34 soil intensified toxicity, suggesting that PAHs-N-PAHs mixtures represent greater risk to soil biota.

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36 **Keywords** Phenanthrene • Nitrogen-containing PAHs • Toxicity • Behavioural index • Soil • Earthworm.

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48 **1 Introduction**

49 Polycyclic aromatic hydrocarbons (PAHs) are a key group of contaminants present at many contaminated sites
50 (e.g. ex-industrial sites, oil contaminated areas) (Brumley et al. 1991; Sverdrup et al. 2002; EC 2011; IARC
51 2012; Anyanwu and Semple 2016b). What is less well described is that their nitrogen-containing polycyclic
52 aromatic hydrocarbons (N-PAHs) can also be present, often at high concentrations (De Voogt and Laane 2009).
53 Many of these compounds are known to be toxic, carcinogenic, mutagenic and genotoxic, thus, are of concern to
54 both biota and human exposure (Bleeker et al. 2002; Sverdrup et al. 2002; Kobetičová et al. 2008; Brar et al.
55 2010; IARC 2012; Anyanwu and Semple 2015a, b, c; 2016a, b). In addition, their toxicity cannot be determined
56 by the number of aromatic rings alone (IARC 2012). N-PAHs contain one or more nitrogen atom(s) in place of
57 carbon atom (Table 1). Due to the substitution of N-atom into one or more of the aromatic rings, and the
58 resulting physico-chemical properties, they differ in terms of persistence, mobility, bioavailability and toxicity
59 compared to analogue PAHs (Sverdrup et al. 2002; Kobetičová et al. 2008; Anyanwu et al. 2013; Anyanwu and
60 Semple 2015a; 2016b). For example, the substitution with N-atom(s) makes N-PAHs more polar, mobile and
61 soluble in the soil environment, and as such, putatively more toxic (Sverdrup et al. 2002; Kobetičová et al. 2008;
62 Anyanwu and Semple 2015 c; 2016a, b).

63 Apart from their environmental occurrence, N-PAHs are discharged into soil from anthropogenic
64 sources such as petroleum related activities, oil spills and combustion processes (Brumley et al. 1991; Webber
65 1994; Švábenský et al. 2009). Previously, N-PAH levels were reported to be 1-10 orders of magnitude lower
66 than those of the homocyclic analogues, however, De Voogt and Laane (2009) measured concentrations ranging
67 from similar to equal to one order of magnitude higher than their homocyclic analogues in sediments.
68 Furthermore, studies have reported that N-PAH chemicals are phyto-toxic, inhibitors to soil microflora and have
69 cellular autolytic impact (Willumsen et al. 2001; 2005; Anyanwu and Semple 2015b; c; 2016a, b). In addition, it
70 has been reported that losses of PAH degradation capacity in contaminated soil may be due to the existence of
71 N-PAHs (Meyer and Steinhart 2000). Irrespective of this, only few studies have investigated N-PAH toxicity on
72 soil invertebrates (Sverdrup et al. 2002; Kobetičová et al. 2008; 2011) and their observed effects ranged from
73 (LC₅₀ (not detected) ND - <2000 mg kg⁻¹ and EC₅₀ 94 - 1033 mg k⁻¹) (Sverdrup et al. 2002; Kobetičová et al.
74 2008; 2011). Although there are some toxicity data for a few selected N-PAH compounds, all the investigations
75 focused on the traditional Organization for Economic Cooperation and Development (OECD) methodologies of
76 percentage survival / effects judgements (OECD, 1984) without considering other criteria, such as the

77 behavioural index (general physical condition) of the organisms during exposure. Monitoring of behavioural
78 index of biota in contaminated soils allows assessment of possible impact of various contaminants on different
79 invertebrate species.

80 Generally, contaminated soils contain complex mixtures of compounds that differ in their physico-
81 chemical properties and toxicity to soil biota. Thus, effects of range of aromatic compounds can be intensified
82 by the presence of other compounds. However, the effect of binary and quinary mixtures of phenanthrene and its
83 N-PAH analogues on soil biota have not been reported in literature. Since information on toxicity is essential for
84 effective assessment of poorly managed chemicals and/or chemical groups for environmental risk assessment,
85 this study therefore investigated the effect of single, binary and quinary mixtures of phenanthrene and its
86 structurally similar nitrogen-containing analogues on mature earthworms (*Eisenia fetida*) in soil using OECD
87 (1984) guidelines (with little deviation) and behavioural index tool.

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89 **2 Materials and methods**

90 2.1 Chemicals

91 Phenanthrene (Phen), 1,10-phenanthroline (1,10-Phen), 1,7-phenanthroline (1,7-Phen), 4,7-phenanthroline (4,7-
92 Phen) and benzo[h]quinoline (B[h]Q) were purchased from Sigma Aldrich Company Ltd, UK (Table 1).

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94 2.2 Test organisms

95 Mature earthworms (*Eisenia fetida*) were purchased from Blades Biological Ltd, UK.

96

97 2.3 Soil preparation

98 Soil (without contamination history) from Myerscough Agricultural College in Lancashire, UK was collected
99 from the top layer of field under pasture (from a depth of approximately 5 - 20 cm). The soil was sandy-loam
100 (19.5% clay, 60.4% sand, 20.0% silt) with an organic matter content of 2.7% and pH 6.5 (Doick et al. 2003).
101 The soil was air dried at room temperature, sieved with 2 mm mesh size, and rehydrated with deionised water
102 back to original water holding capacity (WHC). Spiking procedure followed those described in Doick et al.

103 (2003). One third of soil ($\frac{1}{3}$; 100 g), placed in a bowl, were amended with dissolved chemical standards
104 containing acetone (10 ml) to give concentrations of 10, 100 and 500 mg kg⁻¹. The soils were left to evaporate
105 for 4 hours in the fume hood; after which soils were mixed with the remaining $\frac{2}{3}$ (200 g) soil and amended to
106 80% WHC with deionized water. Control samples were prepared using soil amended with acetone only. After
107 amendment, soils (50 g) were weighed into glass jars and the effects on *E. fetida* was measured for 21 d.
108 Recoveries of tested compounds are at a range of 49.50 ± 0.55% and 104.72 ± 8.00%, except for 1,10-
109 Phenanthroline which was not determined since it could not be detected by GC-MS

110 2.4 Earthworm toxicity assay

111 Determination of the effects of the N-PAHs on soil biota was carried out using the earthworm toxicity assay
112 (OECD, 1984) (with little deviation). Mature *E. fetida* (0.3 – 0.4 g) were selected for the bioassay. Prior to test,
113 the earthworms were depurated for 24 h in petri dishes containing moist filter paper, after which the earthworms
114 were weighed. Soils (50 g) were used for the assay. Three exposure concentrations (10, 100 and 500 mg kg⁻¹)
115 were used for each single, binary and quinary mixture, as well as control samples (0 mg kg⁻¹). Five replicates
116 were used for each concentration. The soils were mixed thoroughly and single earthworm was added to each
117 replicate. Samples were covered with perforated lids and incubated at 12 ± 2°C. Mortality, biomass and
118 behavioural index were the test parameters. Assessment of behavioural index (0 - 2) was carried out after 0, 1, 3,
119 5, 7, 14 and 21 d, with 0 = dead, 1 = moribund and 2 = healthy (Langdon et al. 1999; Anyanwu and Semple
120 2016b). Earthworms were presumed dead if no contractile response was observed when prodded lightly with a
121 blunt probe; moribund when weak contractile was observed; and healthy when sharp and vigorous response was
122 observed (Langdon et al. 1999; Anyanwu and Semple 2016b). The depurated earthworms were weighed before
123 and after exposure to determine changes in weight (biomass). Other physical changes on different parts of the
124 earthworms were also recorded.

125

126 2.5 Statistical analysis

127 The mean weight losses of the earthworms for the replicates were used as a measure of biomass change. The
128 concentration that caused weight loss as compared to control values were calculated on the basis of initial
129 measurement and ANOVA was used to determine the significant impact on biomass. Differences are found to
130 be statistically significant when p<0.05. Further, correlations were determined between the exposure
131 concentrations and changes in biomass using Pearsons correlation. Estimation of the concentration of the

132 chemicals in amended soils that caused 50% mortality (LC_{50}), and the concentration that provoked a response
133 halfway between the baseline and maximum (EC_{50}) (i.e. weight-reduction) during the test period were
134 determined by probit analysis in SPSS 20 software package. Graphs were plotted with SigmaPlot 10.0 version.

135

136 3 Results

137 3.1 Assessment of the aromatic hydrocarbons on the behavioural index of *E. fetida* in soil

138 Apart from the traditional OECD guidelines relating to survival / effect measurements, the study investigated
139 the general health condition of *E. fetida* during exposure. Fig. 1 shows the behavioural indices of *E. fetida*
140 exposed to single, binary and quinary mixtures of phenanthrene and its N-PAH analogues in soil. The 21 d
141 assessment of behavioural index (0 - 2) showed that all the compounds negatively impacted on the health of *E.*
142 *fetida* at the highest concentration of 500 mg kg⁻¹ ($p < 0.05$; $r^2 = 0.982$), with the exception of 1,10-Phen, 4,7-
143 Phen (single mixture) and 1,10-Phen + Phen (binary mixture). Among the N-PAHs, benzo[h]quinoline (single
144 mixture) exhibited greater impact on the general wellbeing of *E. fetida* in soil. For example, it was observed that
145 *E. fetida* became moribund after few hours of exposure to B[h]Q soils and experienced mortality at 3 d in the
146 500 mg kg⁻¹ amendments (Fig. 1).

147 However, 4,7-phenanthroline recorded significant effect on the behavioural index of *E. fetida* in the
148 presence of phenanthrene (binary mixture) at the 500 mg kg⁻¹ amendments ($p < 0.05$) (Fig. 1). In addition,
149 benzo[h]quinoline toxicity was slightly reduced in the presence of phenanthrene because, *E. fetida* experienced
150 moribund phase of 5 d in health prior to mortality. Furthermore, the data showed that quinary mixtures of
151 phenanthrene and N-PAHs significantly affected the behavioural index of the exposed earthworms even at 100
152 mg kg⁻¹ ($p < 0.05$) (Fig. 1). Other observed health effect includes; breakage of clitellum, excretion of yellowish
153 fluid and sores.

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155 3.2 Assessment of the aromatic hydrocarbons on the mortality (LC_{50}) of *E. fetida* in soil

156 During the 21 d exposure, neither mortality nor significant weight losses were observed in the control
157 incubations. The survival (%) of *E. fetida* exposed to single, binary and quinary mixtures of phenanthrene and
158 its nitrogen-containing analogues are shown in Fig. 2. From the result, significant mortality rates were recorded

159 in the 500 mg kg⁻¹ amendments (p<0.05). For example, the concentration–response graph shows that survival
160 (%) of the exposed earthworms ranged from 0% – 20% in soils amended with 500 mg kg⁻¹ chemicals, with the
161 exception of 1,10-Phen, 4,7-Phen (single mixture) and 1,10-Phen + phenanthrene (binary mixture) (Fig. 2). In
162 addition, *E. fetida* suffered high mortality (100%) in the 500 mg kg⁻¹ B[h]Q amendment (Fig. 2a).

163 Furthermore, 100% mortality were observed in the 500 mg kg⁻¹ phenanthrene + 4,7-phenantrolone
164 amendment (binary mixture) (p<0.05). In quinary mixtures however, *E. fetida* recorded 20% – 100% mortality
165 in the 100 mg kg⁻¹ and 500 mg kg⁻¹ amendments, respectively (Fig. 2b). From the result, a trend of increased
166 mortality with increase in concentration was observed compared to the control soil (p<0.05). The toxicity data
167 (LC₅₀) of single, binary and quinary mixtures of phenanthrene and selected N-PAHs on *E. fetida* after 21 d
168 incubation in soil are summarized in Table 2. From the data, the LC₅₀ ranged from ND – 329.3 mg kg⁻¹, with
169 quinary mixture recording the lowest value of 148.4 mg kg⁻¹ (Table 2).

170

171 3.3 Assessment of the aromatic hydrocarbons on the biomass (EC₅₀) of *E. fetida* in soil

172 Impact of the chemicals on the biomass of the exposed earthworms was measured (Table 2). The biomass of
173 dead earthworms was not determined and therefore were assigned zero (0). Figs 3 – 4 shows the weight-changes
174 (biomass) of *E. fetida* exposed to single, binary and quinary mixtures of phenanthrene and its N-PAH analogues
175 in soil. From the result, all the aromatics showed significant biomass-effect on *E. fetida*, and there was a trend of
176 decrease in biomass with increase in chemical concentrations. The concentration–biomass–effect plots recorded
177 notable decline in the weight of the earthworms after 21 d exposure to the aromatic hydrocarbons (Fig. 3).
178 Biomass reductions were observed to be pronounced in the N-PAH amendments, especially 1,7-Phen and B[h]Q
179 (single mixture), and in the binary and quinary mixtures of phenanthrene + N-PAHs (p<0.05) (Figs. 3 – 4).

180 The EC₅₀ measurement of single, binary and quinary mixtures of phenanthrene and its structurally
181 similar N-PAHs on the biomass of *E. fetida* after 21 d incubation are summarized in Table 2. The calculated
182 EC₅₀ values ranged from 13.3 – 148.4 mg kg⁻¹. Among the chemical treatments, B[h]Q recorded the lowest EC₅₀
183 of 17.00 mg kg⁻¹ (Table 2). Analysis of data using ANOVA showed statistically significant difference in the
184 earthworms biomass (weight before exposure and weight after exposure) in all the chemicals, at all the
185 concentrations (p<0.05). Further analysis of data showed high negative correlation between the concentrations
186 and biomass after exposure (p<0.05) in all the chemicals, with the exception of 1,10-Phen and 4,7-Phen which
187 showed medium but non-statistically significant relationships (p>0.05) (Table 2).

188

189 **4 Discussion**

190 Despite the majority of published studies having focused on PAHs and to a lesser extent their heterocyclic
191 analogues, the toxicity data relating to *Eisenia fetida* are still fragmented, with most of the studies investigating
192 single compounds and artificial soil (Sverdrup et al. 2002; Kobetičová et al. 2008; Kobetičová et al. 2011;
193 Anyanwu and Semple 2016b). This study brings to focus information on the behavioural index, survival and
194 biomass-effect of *E. fetida* following exposure to single, binary and quinary mixtures of phenanthrene and its
195 structurally similar N-PAHs.

196 The endpoint of an earthworm toxicity test is normally a measure of mortality; however, mortality is
197 unlikely to be the most sensitive parameter for risk assessment (Langdon et al. 1999; Anyanwu et al. 2013;
198 Anyanwu and Semple 2016b). This current study showed that most of the health effects, such as healthy (2) –
199 moribund (1) – mortality (0) and antagonistic interactions occurred within <24 h – 3 d of exposure in the N-
200 PAHs amended soils with particular reference to benzo[h]quinoline. This pattern of mortality and/or interaction
201 may be attributed more to uptake of the chemical(s) across the dermis rather than through ingestion. This is
202 because, earthworms added to soils amended with 500 mg kg⁻¹ N-PAHs were initially healthy and very active,
203 but became moribund after a few hours and died within 3 d. This phenomenon has also been reported for the
204 highest concentration of metal and aromatic hydrocarbon amended soils (Spurgeon et al. 1994; Anyanwu and
205 Semple 2016b). Presumably, as earthworm moves through the soil, they are exposed to N-PAHs, which may
206 lead to absorption of the chemicals across the dermis. This further showed that earthworms were particularly
207 sensitive to N-PAHs compared to homocyclic phenanthrene analogue. Furthermore, other behavioral response
208 such as lack of burrowing into the amended soil and avoidance (moving away from the amended soil) was
209 observed at the highest concentration (500 mg kg⁻¹).

210 In this current study, benzo[h]quinoline was noted to be the most toxic chemical to *E. fetida*. It may be
211 that N-PAHs such as benzo[h]quinoline, having log K_{ow} of 3.43 and log K_{oc} of 4.32, may be existing more in the
212 aqueous phase (even though some may be associated with soil organic matter), hence, undergoing rapid
213 absorption and thus, triggering physical injury to the dermis of the earthworms as observed by the formation of
214 lesions, breakage of the clitellum, sores, loss of weight and eventually death (Broholm et al. 1999; Anyanwu and
215 Semple 2015b).

216 Variations in aromatics toxicity were observed. The variations in toxicity could be ascribed to the
217 differences in physico-chemical properties and N-position (Anyanwu et al. 2013; Anyanwu and Semple 2015a,
218 b, c; 2016 a, b). Furthermore, variability was measured among 1,7-phen, 1,10-phen and 4,7-phen irrespective of
219 their similarities in physico-chemical properties. Molecular structure and/or chemical bioavailability may be
220 attributed in this study. From the results, the recorded LC₅₀ and EC₅₀ values for phenanthrene are in agreement
221 with the values reported by Sverdrup et al. (2002) (Table 3). However, the values (LC₅₀ and EC₅₀) were lower
222 than those reported by Kobetičová et al. (2008; 2011); soil physico-chemical properties and/or species
223 differences may be attributable. Further, the absence of mortality recorded in the 1,10-Phen amendment (in the
224 study) is in agreement with the findings of Sochová et al. (2007), who reported 1,10-Phen as the least toxic
225 chemical to nematode (*Caenorhabditis elegans*) in soil. However, the observation was different from the reports
226 of Kobetičová et al. (2008; 2011), who recorded high toxicity for 1,10-Phen; chemical concentrations and/or
227 media (type of matrix) may be ascribed. In support, Anderson et al. (1999) reported that during chemical-biota-
228 interactions, the receptor, route of entry, test duration and the matrix containing the contaminant need to be
229 considered. Therefore, with a soil organic matter content of 2.7% and pH 6.5, the toxicity and speciation of
230 1,10-Phen may have been influenced (in this study). Similarly, (although not in the soil environment) the
231 observation with 4,7-Phen (single mixture) is in agreement with Feldmannová et al. (2006), who recorded 4,7-
232 Phen as the least toxic compound on the survival, fecundity and reproduction of *Daphnia magna*.

233 It has been reported that biomass of organisms (such as earthworms) can be impaired by: (i) direct
234 toxic effects on the physiology of exposed organism, (ii) changes in the body function as the organism tries to
235 prevent accumulation in the biological membranes and/or (iii) avoidance due to lack of feeding. Thus, the
236 significant effect on biomass (weight) of *E. fetida* recorded in this current study may be ascribed to toxicity
237 and/or changes in body function. However, it should be noted that the toxicological effects of N-PAHs could
238 cause reductions in weight (Sverdrup et al. 2002; Anyanwu et al. 2013; Anyanwu and Semple 2016b). In
239 addition, Widdows and Donkin (1989) reported changes in the individual energy budget as an organism (such as
240 earthworm) spends energy resisting the contaminant by avoidance, elimination or reluctant to feed in a polluted
241 environment. According to their report, the extra energy requirement will decrease the capacity for growth and
242 development of the organism. This suggests that the effect on biomass of the exposed earthworms is as a result
243 of changes in the energy budget as they strive to resist bioaccumulation in the site of biological response, or
244 weight loss through lack of feeding and/or N-PAHs toxicity. Furthermore, the study showed that the biomass–

245 effect ratios increased with increase in N-PAHs burden on the earthworms as demonstrated by the correlation
246 relationships; indicating greater toxicity of N-PAHs to soil biota.

247

248 **5 Conclusions**

249 In this study, the effects of single, binary and quinary mixtures of phenanthrene and its structurally similar N-
250 PAHs were investigated since mixtures of contaminants are found in contaminated soils. The results showed
251 that single N-PAHs possess high ecotoxicity risk as expected of their solubility and lower K_{ow} values, while
252 mixtures of phenanthrene and N-PAHs in contaminated sites represent greater risks to soil biota. However,
253 studies are required on the impact of N-PAHs in chronically contaminated / aged soil.

254

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258

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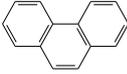
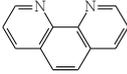
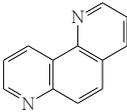
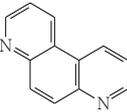
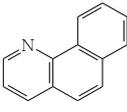
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352 **Table 1** Test chemicals and their physico-chemical properties

Chemical	Chemical formula	Chemical structure	Molecular mass	Boiling point (°C)	Log K_{ow}	Solubility 25°C (mg L ⁻¹)	% purity
Phenanthrene	C ₁₄ H ₁₀		178.20	340.00	4.46	1.15	96.00
1,10-Phenanthroline	C ₁₂ H ₈ N ₂		180.21	365.10	2.51	30.64	99.00
1,7-Phenanthroline	C ₁₂ H ₈ N ₂		180.21	365.10	2.51	30.64	99.00
4,7-Phenanthroline	C ₁₂ H ₈ N ₂		180.21	361.20	2.40	38.04	98.00
Benzo[h]-quinoline	C ₁₂ H ₉ N		179.20	339.00	3.43	78.70	97.00

353 Source: www.chemspider.com/Chemical-Structure, Anyanwu and Semple (2015a, b, c ; 2016 a, b)

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367 **Table 2** Summary of the effect of single, binary and quinary mixtures of phenanthrene and its N-PAH analogues
 368 on the survival (LC₅₀) and biomass (EC₅₀) of *E. fetida* after 21 d incubation in soil

Chemical	LC ₅₀ (mg kg ⁻¹)	EC ₅₀ (mg kg ⁻¹)	R ² (biomass)
Phen	329.30 (ND)	148.40 (ND)	-0.976
1,10-Phen	ND	102.30 (ND)	-0.686
1,7-Phen	219.80 (ND)	93.90 (ND)	-1.00
4,7-Phen	ND	13.30 (ND)	-0.690
B[h]Q	219.80 (ND)	17.00 (0-88.10)	-0.972
1,7-Phen + Phen	219.80 (ND)	63.80 (ND)	-0.979
4,7-Phen + Phen	219.80 (ND)	148.40 (ND)	-0.956
B[h]Q + Phen	219.80 (ND)	63.80 (5.5-526.70)	-0.979
Phen + N-PAHs	148.40 (ND)	24.20 (ND)	-0.961

369 Values show LC₅₀, EC₅₀ and 95% confidence interval (in parenthesis), ND = (not determined), n = 5.

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387 **Table 3** LC₅₀ and EC₅₀ ranges of phenanthrene and its nitrogen-containing analogues on survival and growth of
 388 soil organisms

Chemical	Test organism	Test duration	LC ₅₀ (mg kg ⁻¹)	EC ₅₀ (mg kg ⁻¹)	Reference
	<i>E. veneta</i>	28 d	134 (ND)	94 (64-125)	Sverdrup <i>et al.</i> 2002
Phen	<i>E. cryptius</i>	28 d	1708 (1494-1920)	869 (627-1110)	Kobetičová <i>et al.</i> 2011
	<i>E. fetida</i>	21 d	329.3 (ND)	148.4 (ND)	This study
1,10-Phen	<i>E. fetida</i>	4 wks	1500<LC ₅₀ <2000	1033 (986-1097)	Kobetičová <i>et al.</i> 2008
	<i>E. cryptius</i>	28 d	ND	798 (653-939)	Kobetičová <i>et al.</i> 2011
	<i>E. fetida</i>	21 d	ND	102.3 (ND)	This study
1,7-Phen	<i>E. fetida</i>	21 d	219.8 (ND)	93.9 (ND)	This study
4,7-Phen	<i>E. fetida</i>	21 d	ND	13.3 (ND)	This study
B[h]Q	<i>E. fetida</i>	21 d	219.8 (ND)	17.0 (0-88.1)	This study

389 Values show LC₅₀, EC₅₀ and 95% confidence interval (in parenthesis), ND = (not determined).

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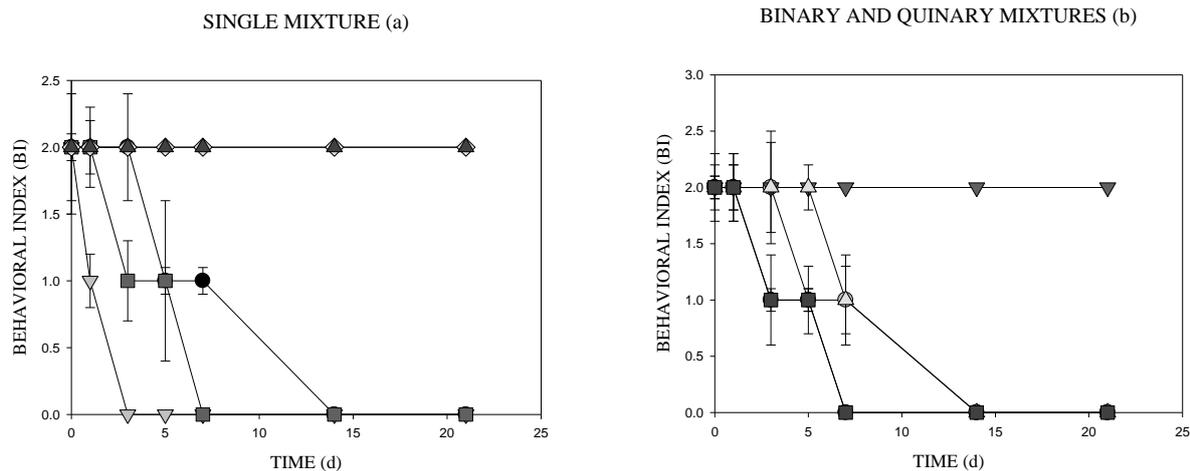
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401 **Fig. 1** Behavioral index of *E. fetida* exposed to 500 mg kg⁻¹ of single, binary and quinary mixtures of
 402 phenanthrene and its N-PAHs analogues in soil during 21 d incubation. Data shows: Phen (●); B[h]Q (▼); 1,7-
 403 Phen (■); 1,10 (◇) and 4,7-Phen (▲) (single mixture) (**Fig. 1a**). B[h]Q + Phen (●); 1,7-Phen + Phen (○); 1,10-
 404 Phen + Phen (▼); 4,7-Phen + Phen (▲) and Phen + NPAHs (■) (binary and quinary mixtures) (**Fig. 1b**).

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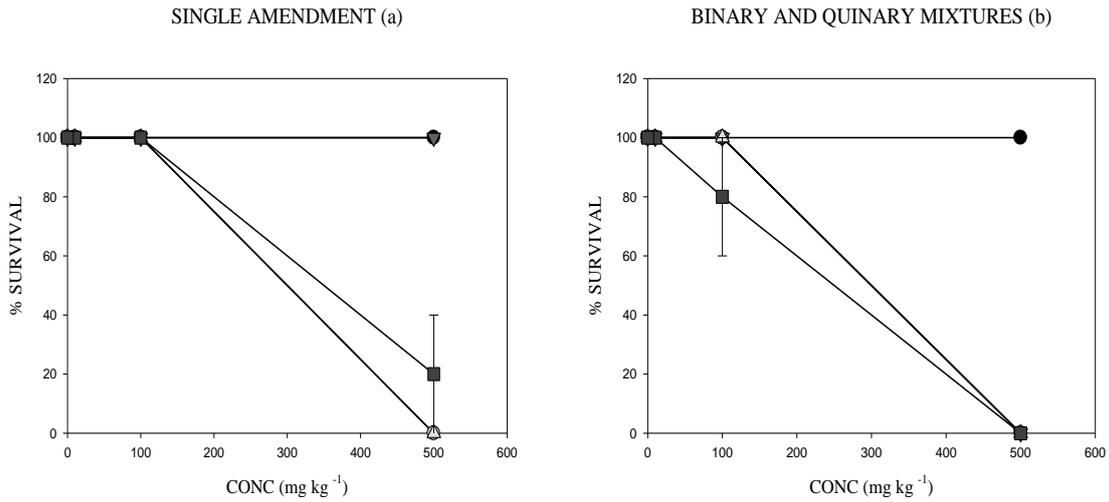
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422 **Fig. 2** Survival (%) of *E. fetida* exposed to single, binary and quinary mixtures of phenanthrene and its N-PAH
 423 analogues in soil at 0 (control), 10, 100 and 500 mg kg⁻¹ after 21 d incubation. Data shows: 1,10-Phen(●); 1,7-
 424 Phen (○); 4,7-Phen (▼); B[h]Q (Δ) and Phen (■) (single amendment) (**Fig. 2a**). 1,10-Phen + Phen (●); 1,7-Phen
 425 + Phen (○); 4,7-Phen + Phen (▼); B[h]Q + Phen (Δ) and Phen + NPAHs (■) (binary and ternary mixtures) (**Fig.**
 426 **2b**).

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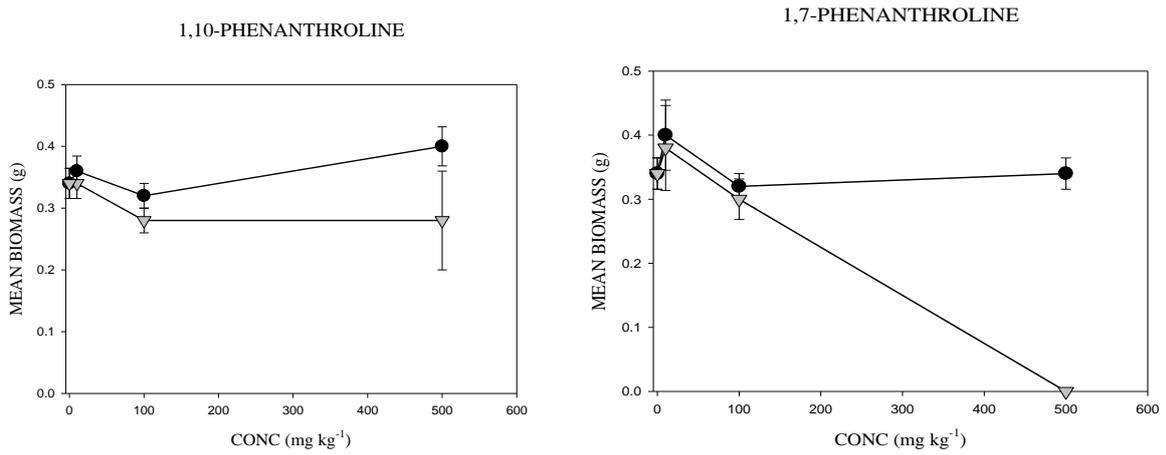
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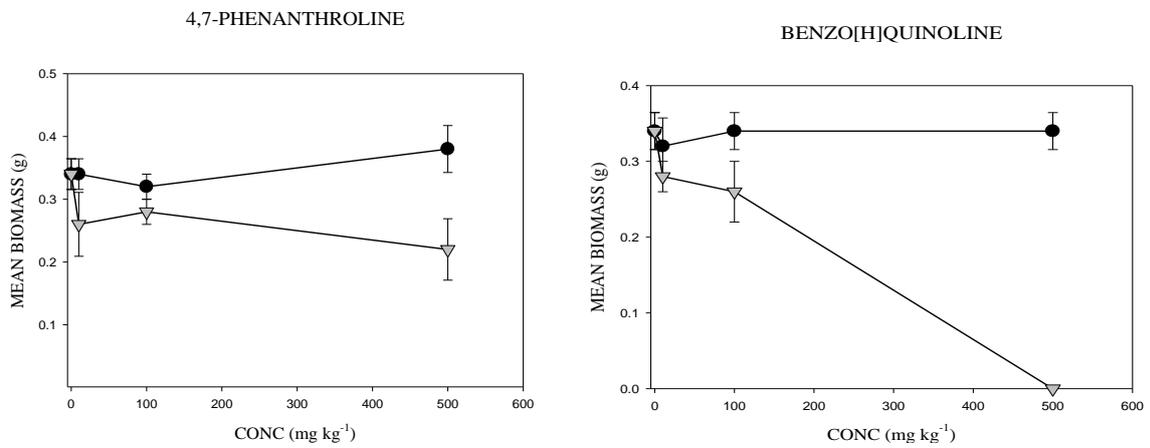
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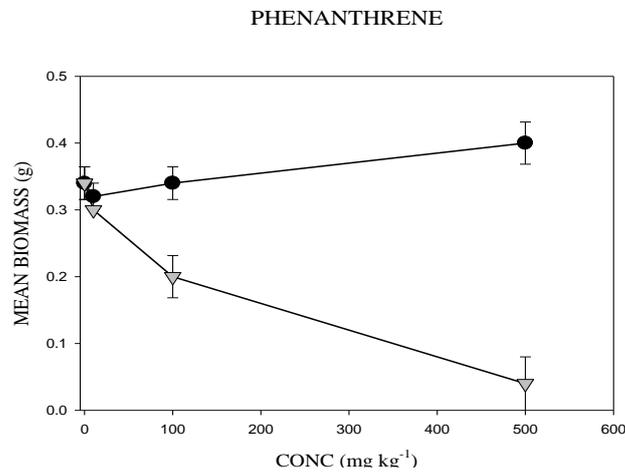
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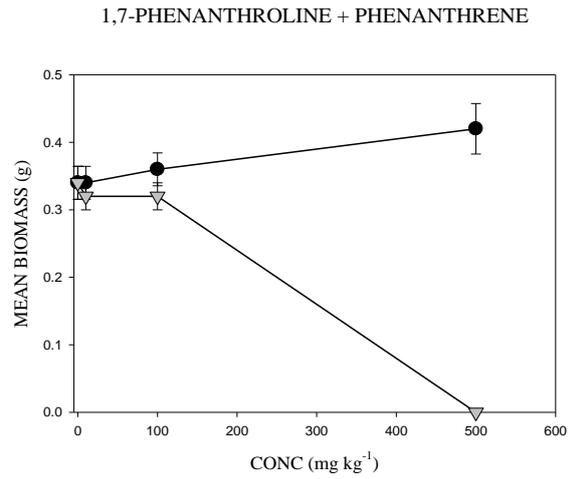
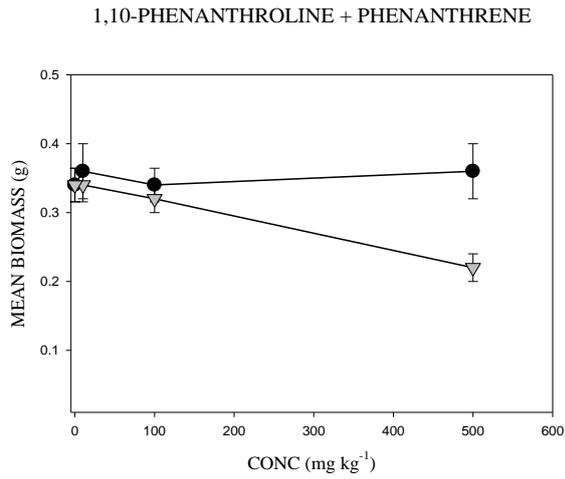


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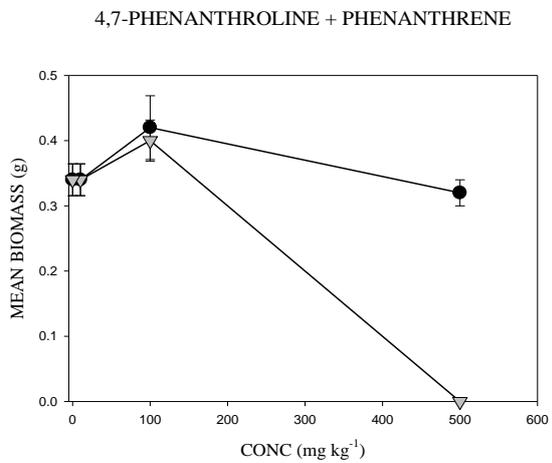


441 **Fig. 3** Mean biomass (g) of *E. fetida* exposed to single amendments of phenanthrene and its N-PAH analogues
442 in soil at 0 (control), 10, 100 and 500 mg kg⁻¹ after 21 d incubation. Data shows: weight before exposure (●) and
443 weight after exposure (▼).

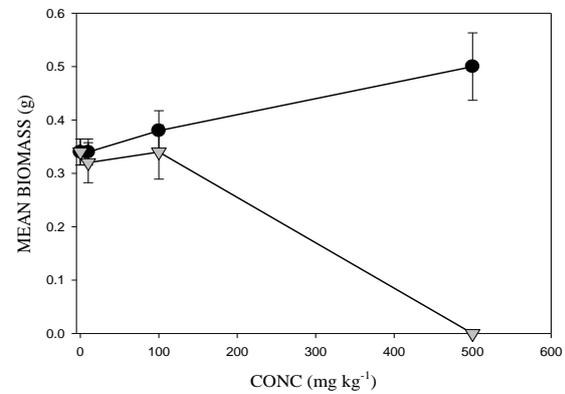
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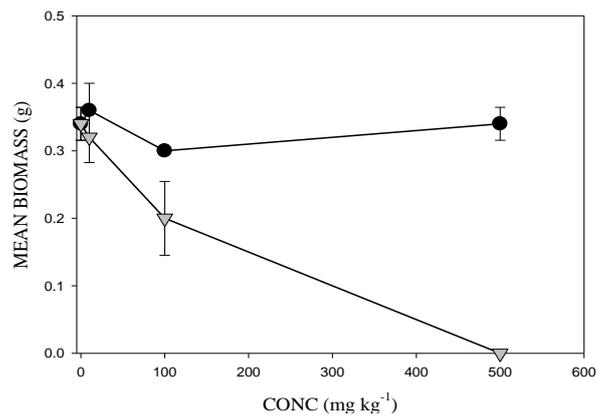


BENZO[H]QUINOLINE + PHENANTHRENE



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PHENANTHRENE + N-PAHs



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448 **Fig. 4** Mean biomass (g) of *E. fetida* exposed to binary and quinary mixtures of phenanthrene and its N-PAH
 449 analogues in soil at 0 (control), 10, 100 and 500 mg kg⁻¹ after 21 d incubation. Data shows: weight before
 450 exposure (●) and weight after exposure (▼).